Interaction of Hexacyanoferrate(I1) with Cyanocobalamin. Kinetics and Reaction Mechanisms

YOUSIF SULFAB and MOHAMED S. EL-EZABY* *Chemistry Department, Faculty of Science, Kuwait University, Kuwait* Received April 27, 1977

In acidic media, the kinetically stable cyanocobalamin was observed to undergo rapid substitution in accordance to the rate law

$$
k_{obs} = a + bT_{Fe}
$$

with a and b correlated to the backward and forward processes, respectively. The mechanism of substitution is discussed. The equilibrium constant for the acid dependent path is 9.3 X IO3 and for the independent path is 4.75 X 102.

Introduction

Various aspects of the biochemical action of cobalt corrinoids have been reported [1, 2]. Surprisingly, not a great deal of work has been published on the kinetics of ligand substitution in cobalamins. Few works have investigated the displacement of the water molecule in aquacobalamin (3-S) by the ligands azide, cyanate, imidazole, thiocyanate, sulfite, thiosulphate, iodide and bromide. These reactions are rapid in contrast to the fact that octahedral Co(II1) complexes are inert. Thusius [5], in contrast to Randall and Alberty $[3, 4]$, has concluded that the dissociation of water in the *trans* position in aqua-cobalamin is the rate determining step followed by rapid addition of the entering ligand.

It has been also known that the ligand trans to a u-bonded alkyl substituent undergoes substitution at rates considerably higher than the same Iigand in similar compounds lacking M-C bond [6, and references therein]. The kinetic expressions which have been exposed to examination (in such cases) take one of the following forms:

Scheme I. A limiting S_N1 mechanism

$$
R(Co)H_2O \xrightarrow[k-1]{k_1} R(Co) + H_2O
$$

$$
R(Co) + L \frac{k_2}{k_{-2}} R(Co)L
$$

$$
k_{obs} = \frac{(k_1 k_2 / k_{-1})(L) + k_{-2}}{1 + (k_2 / k_{-1})(L)} (L \gg R(Co)H_2O) (1)
$$

Scheme II. Outer-sphere complexation mechanism

$$
R(Co)H_2O L \xrightarrow{K_{IP}} R(Co)H_2O \cdots L
$$

\n
$$
R(Co)H_2O \cdots L \xrightarrow[K_{-3}]{k_3} R(Co)L + H_2O
$$

\n
$$
k_{obs} = \frac{(k_3 + k_{-3}) K_{iP}(L) + k_{-3}}{1 + K_{iP}(L)} (L \gg R(Co)H_2O)
$$
 (2)

In fact, reported experimental kinetic data for the equilibrium

$$
R(Co)H2O + L = R(Co) L + H2O
$$

is of the form

$$
k_{obs} = k_f(L) + k_b
$$
 (3)

Comparing eq. (3) with eqs. (1) and (2) indicates that either $(k_2/k_{\text{t}}) L \ll 1$ in scheme 1 or $K_{ip}(L) \ll 1$ 1 in scheme II.

In the present work we report the results of the kinetics of interaction of hexacyanoferrate(I1) with cyanocobalamin in acidic solution.

Experimental

Reagents

Analytical grade $K_4Fe(CN)_6.3H_2O$ and Vitamin B_{12} (cyanocobalamin) were used without further purification. A stock solution of hexacyanoferrate(I1) $(10, F)$ was freshly prepared in all kinetic runs. ock solution of 1.4×10^{-3} *F* evanocobalamin was prepared and kept in a dark container.

Kinetic runs were carried out using a Durrum/ Gibson Stopped-Flow apparatus thermostatically con-

^{*}To whom correspondence should be addressed.

Figure 1. Spectra of cyanocobalamin at different acid concentrations.

rolled at 25 °C . Optical path-length was 2.0 cm . The wavelength used was 550 nm.

The spectrum of vitamin B_{12} in presence and absence of hexacyanoferrate(I1) at different total acid concentrations were recorded on Sp 8000 PYE-UNICAM double beam spectrophotometers. The concentration range of cyanocobalamine was (2.95-5.90) 10^{-5} *F* and that of hexacyanoferrates was $(0.4-2.0)$ 10^{-3} *F*. The total hydrochloric acid concentration was in the range $0.1-1$ *F*. Ionic strength was kept at constant value of 1.0 *F* using the appropriate mixtures of HCl and KC1 solutions.

Results and Discussion

A change in color (pink to orange pink) has been observed when cyanocobalamin was acidified with HCl solution. The spectra of cyanocobalamin solutions at different hydrogen ion concentrations are shown in Fig. 1. The bands are slightly shifted to shorter wavelengths as hydrogen ion concentrations increases. When hexacyanoferrate(I1) solution was added to the acidified solution of cyanocobalamin a rapid change to the original color has been observed.

Earlier work had indicated that cyanocobalamin is protonated in acidic media **[l] .** The nitrogen atom of α -5,6-dimethylbenzimidazole nucleotide group *trans* to the cyano ligand is readily protonated in acidic medium with its position being occupied

Figure 2. (a) The absorbance of cyanocobalamin as function of T_H at λ 550 nm and μ = 1.0 *F* at 25 °C. (b) The plot of $(T_VT_H/1\epsilon_2T_V - As_i)$ as function of T_H .

by a water molecule or chloride ion. The protonation equilibrium constant was reported to be ~ 0.8 [1]. Other protonation constants are of very small magnitude. This value has been checked further in this work. The spectra of cyanocobalamin were taken at different hydrochloric acid concentrations. The absorbance at 550 nm was taken as function of the total acid concentrations (T_H) shown in Fig. 2(a). The protonation constant may be calculated from the following equation

$$
\frac{(H)_i T_v}{1 \epsilon_2 T_v - As_i} = \frac{1}{K_v \Delta \epsilon} + \frac{(H)}{\Delta \epsilon}
$$
 (5)

where T_v is the total concentration of cyanocobalamin; As_i the absorbance at a given wavelength; $\Delta \epsilon$ = $\epsilon_2 - \epsilon_1$ where ϵ_1 and ϵ_2 are the molar absorptivities of the protonated and nonprotonated species of cyanocobalamin and K_v the protonation constant.

The molar absorptivity ϵ_2 was taken from the absorbance value of cyanocobalamin only in 1 M KCl, and (H) was assumed to be equal to the total acid concentration. By plotting the left hand-side term versus T_H a linear plot has been obtained with intercept and slope equal to $(1.13 \pm 0.07) 10^{-4}$ and $(1.18 \pm 0.10) 10^{-4}$, respectively, Fig. 2(b). The calculated value of K_v amounts to 1.04 \pm 0.11, which is not different significantly from the reported value of ≈ 0.80 .

On the other hand, numerous protonated species of hexacyanoferrate(I1) coexist in solution. In fact, 5 species may exist as the following equilibria indicate

Figure *3.* The dependence of observed rate constant on both acid concentration and hexacyanoferrate(I1).

a)
$$
Fe(CN)_{6}^{4-} + H^{+} \longrightarrow HFe(CN)_{6}^{3-};
$$

$$
K_{A} = \frac{[HFe(CN)_{6}^{3-}]}{[H^{+}][Fe(CN)_{6}^{4-}]}
$$

b)
$$
HFe(CN)_{6}^{3-} + H^{+} \longrightarrow H_{2}Fe(CN)_{6}^{2-};
$$

$$
K_{B} = \frac{[H_{2}Fe(CN)_{6}^{2-}]}{[HFe(CN)_{6}^{3-}][H^{+}]}
$$

c)
$$
H_2Fe(CN)^{2-}_{6} + H^+ \longrightarrow H_3Fe(CN)^{-}_{6};
$$
 (6)

$$
V = [H_3Fe(CN)^{-}_{6}]
$$

$$
{}^{\mathbf{R}_{\mathbf{C}}} = \left[H_2 \mathrm{Fe(CN)}_{6}^{2-} \right] \left[H^{\dagger} \right]
$$

d)
$$
H_3Fe(CN)^{-}_{6} + H^{\dagger} \longrightarrow H_4Fe(CN)^{0}_{6};
$$

$$
K_D = \frac{[H_4Fe(CN)_6^2]}{[H_3Fe(CN)_6][H^4]}
$$

 $W(x,y)$

Few reports studied the above equilibria [7], however only K_A and K_B have been determined. The values btained for K, and K_n are 5.2×10^2 and 0.83 X 0^2 respectively. The third and forth equilibria are expected to have constants of very low values (≤ 0.1) .

The kinetic study was carried out at the wavelength 550 nm in the acid concentration range of 0.1-1.0 *F.* Figure 3 shows the dependence of observed rate constant, k_{obs} , on both T_H and total concentration of hexacyanoferrate, T_{Fe} . At a given acid (T_H) concentration, k_{obs} is linearly dependent on T_{Fe} (Fig. 3) *i.e.,*

$$
k_{\rm obs} = a + bT_{\rm Fe}
$$
 (7)

The values of a and b are listed in Table I. Interaction of hexacyanoferrate(I1) species with that of cyanocobalamin may follow scheme III.

At a given hydrogen ion concentration, the rate equation describing the mechanism in scheme(II1) can be expressed as follows:

TABLE 1. The Values of the Constants a and b at Different Acid Concentrations.

${\bf T_H}$, F	a, \sec^{-1}	$b \times 10^{-7}$, sec ⁻¹ M^{-1}	R
0.10	51.2 ± 6.5	3.24 ± 0.49	0.978
0.25	53.4 ± 6.6	4.38 ± 0.48	0.988
0.50	56.4 ± 4.6	5.43 ± 0.49	0.988
1.00	63.2 ± 5.2	7.24 ± 0.44	0.996

$$
-\frac{d[(I) + (II) + (III) + (IV) + (V)]}{dt} = -Z \frac{d(V)}{dt} =
$$

= $\frac{T_{Fe}(V)}{X} \left[\frac{(H^4)U}{K_V} + W \right] - (V)(H^4)M + N$ (8)

where

$$
X = (H^{\dagger})^4 + (H^{\dagger})^3 K_A + (H^{\dagger})^2 K_A K_B + (H^{\dagger}) K_A K_B K_C + K_A K_B K_C K_D,
$$

$$
U = k_1(H^{\dagger})^4 + k_2(H^{\dagger})^3 K_A + k_3(H^{\dagger})^2 K_A K_B + k_4(H^{\dagger}) K_A K_B K_C + K_A K_B K_C K_D,
$$

$$
W = k_6 (H^{'})^4 + k_7 (H^{'})^3 K_A + k_8 (H^{'})^2 K_A K_B + k_9 (H^{'}) K_A K_B K_C + K_{10} K_A K_B K_C K_D,
$$

$$
M = k_{-1} \frac{(H')^{4}}{Q_{1}} + k_{-2} \frac{(H')^{3}}{Q_{II}} + k_{-3} \frac{(H')^{2}}{Q_{III}} + k_{-4} \frac{(H')^{4}}{Q_{IV}} + k_{-5}
$$

$$
N = k_{-6} \frac{(H')^{4}}{Q_{I}} + k_{-7} \frac{(H')^{3}}{Q_{II}} + k_{-8} \frac{(H')^{2}}{Q_{III}} + k_{-9} \frac{(H')^{4}}{Q_{IV}} + k_{-10}
$$

$$
Z = \frac{(H')^{4}}{Q_{I}} + \frac{(H')^{3}}{Q_{II}} + \frac{(H')^{2}}{Q_{III}} + \frac{(H')^{2}}{Q_{IV}} + I
$$

where k_i 's are the forward rate constant, k_i 's are the backward rate constants, I, II, III, IV, V are the prouct species, and Q_2 , Q_{II} , Q_{II} , Q_{II} , are the overall $\frac{1}{2}$ enrotonation constants of I, II, III, IV species. The vertical protolytic reactions of the above scheme are assumed to be at equilibrium since they are fast in comparison with the complex formation reactions. Since the total acid concentration is not altered during the reaction progress, the integrated expression of eq. 9 can be obtained

$$
\ln \frac{(V)_{eq}}{(V)_{eq} - (V)_t} = k_{obs}t
$$
 (9)

Figure 4. The dependence of a on hydrogen ion concentration.

where $(V)_{eq}$ and $(V)_{t}$ are the concentrations of the complex at equilibrium and at time t and

$$
k_{\rm obs} = \left[\left(\frac{(H^{\dagger})U}{X K_V} + \frac{W}{X} \right) \middle/ \left(1 + \frac{(H^{\dagger})}{K_V} \right) \right] T_{\rm Fe} + \frac{(H^{\dagger})M + N}{Z}
$$

The observed rate constant, k_{obs} , was obtained experimentally from the slope of the plot of $ln(As_{eq}/)$ $\text{As}_{\text{eq}} - \text{As}_{t}$) vs. t when $T_{Fe} \gg T_{V}$. The derived rate constant can be correlated with that obtained experimentally (eq. (7)), *i.e.*

$$
a = \frac{(H^*)M + N}{Z} \tag{10}
$$

$$
b = \left[\frac{(H^*)U}{XK_V} + \frac{W}{X}\right] / \frac{1 + (H^*)}{K_V}
$$
 (11)

The validity of eq. 8 and the linear dependence of k_{obs} on T_{Fe} indicate that the rate of complex reaction is first order with respect to cyanocobalamin and hexacyanoferrate concentration. Moreover the linear dependence (Fig. 4) of 'a' on hydrogen ion concentration indicates that Z is practically equal to one. From the slope and intercept of the straight line, the M and N values were obtained $(M = 13.2 \pm 0.2$ and $N = 49.9 \pm 0.1$.

The foregoing conclusion indicates further that the terms

$$
k_{-1} \frac{(H')^4}{Q_1}
$$
, $k_{-2} \frac{(H')^3}{Q_{II}}$, $k_{-3} \frac{(H')^2}{Q_{III}}$ and $k_{-4} \frac{(H')}{Q_{IV}}$

may be ignored with respect to k_{-5} and the terms

$$
k_{-6}
$$
 $\frac{(H')^4}{Q_I}$, k_{-7} $\frac{(H')^3}{Q_{II}}$, k_{-8} $\frac{(H')^2}{Q_{III}}$, k_{-9} $\frac{(H')}{Q_{IV}}$

Figure 5. The dependence of $b(1 + T_H/K_V)$ on hydrogen ion concentration.

may also be ignored with respect to k_{-10} . On the other hand, the dependence of b on hydrogen ion concentration looks complicated as eqn. 10 indicates. However, little rearrangement may show the dependence

$$
b\left[1 + \frac{(H^*)}{K_V}\right] = \frac{U}{XK_V}(H^*) + \frac{W}{X}
$$
 (12)

The plot of the left-side term of eqn. 12 versus $(H⁺)$ showed a linear dependence, Fig. 5, with intercept (= $W/X = (2.37 \pm 0.15) \times 10^4$) and slope (= U/ $X_{\text{IV}} = (11.80 \pm 0.26) 10^{4}$. This conclusion indicated that the terms U/XK_V and W/X are independent of hydrogen ion concentration. In other words, U/XK_V should be reduced to k_5/K_V from which k_5 can be calculated by knowing K_V , *i.e.*

$$
k_5 = (1.23 \pm 0.13)10^5
$$
 and $\frac{W}{X} = k_{10}$

From the foregoing results, one can reach the conclusion that reactions e and j are quite significant in the above reaction mechanism. In scheme (III), we have not accounted for the rapid equilibrium which may exist between pentacoordinate and hexacoordinate species of cyanocobalamin or base-off and base-on forms [8]. If these forms are taken in consideration two routes may be distinguished:

Scheme (III)

a) H₄Fe(CN)₆ + CN·Co·BM·H⁺
$$
\xrightarrow[k-1]
$$
 CN·Co·BM(H₄Fe(CN)₆ + H⁺
\nb) H₃Fe(CN)₆ + CN·Co·BM·H⁺ $\xrightarrow[k-2]$ CN·Co·BM(H³Fe(CN)₆ + H
\nc) H₂Fe(CN)₆²⁺ + CN·Co·BM·H⁺ $\xrightarrow[k-2]$ CN·Co·BM(H₂Fe(CN)₆²⁺ + H
\n1) H⁺
\n2) H₅Fe(CN)₆²⁺ + CN·Co·BM·H⁺ $\xrightarrow[k-3]$ CN·Co·BM(H₂Fe(CN)₆²⁺ + H
\n1) H⁺
\n3) H₆(CN)³₆ + CN·Co·BM·H⁺ $\xrightarrow[k-4]$ CN·Co·BM(H₇Fe(CN)₆³⁺ + H
\n2) F₆(CN)⁴₆ + CN·Co·BM·H⁺ $\xrightarrow[k-5]$ CN·Co·BM(F₆(CN)₆⁴⁻ + H
\n3) H₄Fe(CN)₆ + CN·Co·BM $\xrightarrow[k-5]$ CN·Co·BM(H₄Fe(CN)₆)
\ng) H₃Fe(CN)₆ + CN·Co·BM $\xrightarrow[k-5]$ CN·Co·BM(H₃Fe(CN)₆)
\nh) H₂Fe(CN)₆²⁻ + CN·Co·BM $\xrightarrow[k-5]$ CN·Co·BM(H₂Fe(CN)₆)
\ni) H₆(CN)₆²⁺ + CN·Co·BM $\xrightarrow[k-5]$ CN·Co·BM(H₇Fe(CN)₆

One route in which species (I) and (II) react with hexacyanoferrate(II) species. Another route involves the reaction of species (III) and (IV) with the hexacyanoferrate(II) species. Since the observed rate was only detected in acidic solution one may exclude the former route and substitution by the hexacyanoferrate(II) species should involve species (III) and (IV). In such case, it is not simple to differentiate between a limiting S_N1 mechanism or outer-sphere complexation mechanism.

If our kinetic conclusions are correct the equilibrium constants of reactions e and j in scheme (III) can be calculated. The values so obtained are K_e = k_5/k_{-5} = (9.30 ± 0.99) 10³ and K_i = k₁₀/k₋₁₀ = $(4.75 \pm 0.30) 10^2$.

The values of K_e and K_i are expected to be of equal magnitude since $K_v = 1.04$, but $K_e/K_i = 20$. This difference can be seen as resulting from the formation of an outer-sphere complex in reaction (e)

in scheme III. The reaction path j in scheme III is probably dissociative.

References

- 1 J. M. Pratt, "Inorganic Chemistry of Vitamin B₁₂", Academic Press, New York (1972).
- 2 D. G. Brown, in "Current Research Topics in Bioinorganic Chemistry", Progress in Inorganic Chemistry, Vol. 18, Ed. by S. J. Lippard, p. 177 (1973).
- 3 W. C. Randall and R. A. Alberty, Biochem., 5, 3189 $(1966).$
- 4 W. C. Randall and R. A. Alberty, Biochem., 6, 1520 $(1967).$
- 5 D. Thusius, J. Am. Chem. Soc., 93, 2629 (1971).
- 6 J. H. Espenson and R. Russell, 13, 1, 7 (1974).
- 7 J. Jordan and G. J. Ewing, *Inorg. Chem.*, 1, 587 (1962).
8 D. G. Brown, "The Chemistry of Vitamin B₁₂ and Related
- Inorganic Model Systems", in Progress in Inorganic Chemistry, Vol. 18, Ed. by S. J. Lippard, p. 177, Wiley, N.Y. $(1973).$